





#### **Berl R. Oakley**, Tomohiro Akashi and C. Elizabeth Oakley

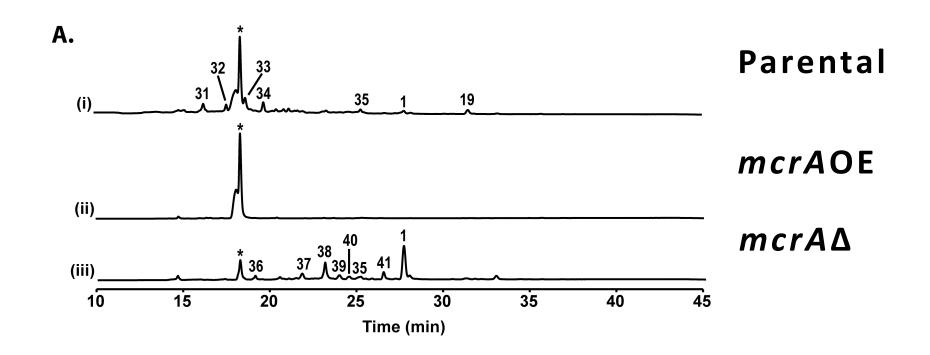


#### What is McrA (AN8694)?

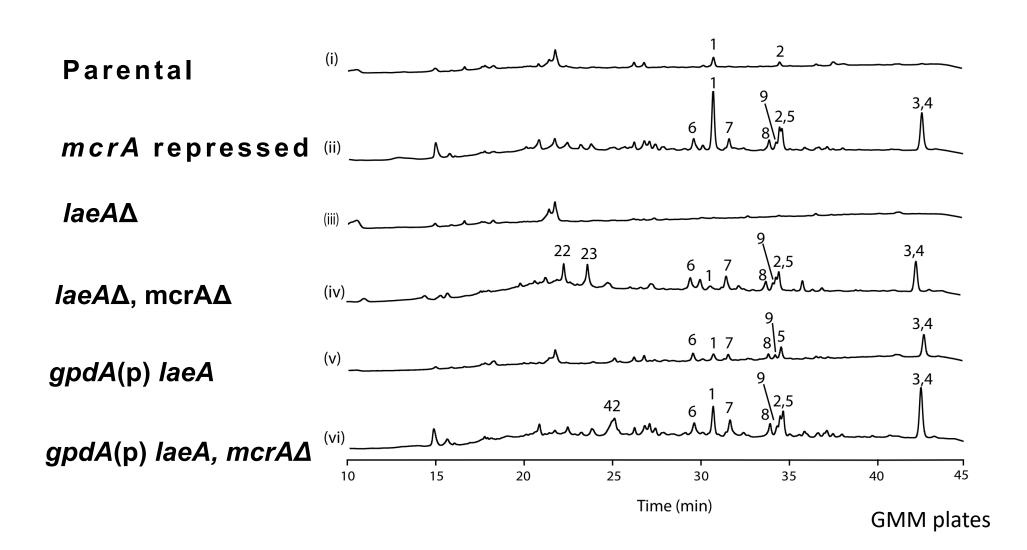
- It was identified in a screen for negative regulators of secondary metabolism in *Aspergillus nidulans* (Oakley et al., 2017, Molecular Microbiology 103:347-365).
- It is strongly predicted to be a transcription regulator with a zincfinger DNA binding domain.
- It is conserved among ascomycetes (1 or 2 strong homologs/genome).



McrA regulates production of many secondary metabolites









- McrA plays a role in the regulation of at least 10 secondary metabolite gene clusters.
- Most are upregulated in the *mcrA* deletion.
- Deletions of *mcrA* homologs in *Aspergillus terreus* and *Penicillium canescens* also upregulate secondary metabolite production.
- Production of compounds in *mcrA*∆ strains is influenced by media (carbon source) and growth conditions.



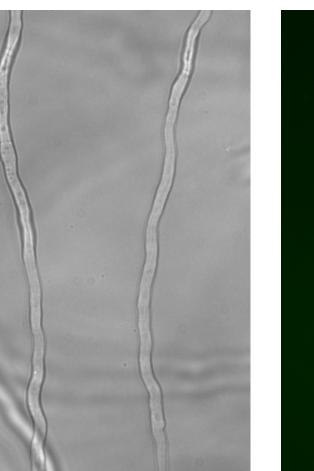
### Effects of mcrA Deletion on transcription

- Deletion of *mcrA* significantly alters the transcription of 1352 genes (p<0.05).
- Genes related to primary and secondary metabolism are enriched among transcriptionally altered strains.
- The majority of transcriptionally altered genes are upregulated.

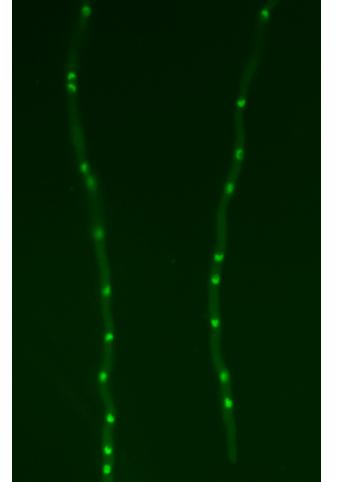


#### McrA-GFP localizes to the nucleoplasm.

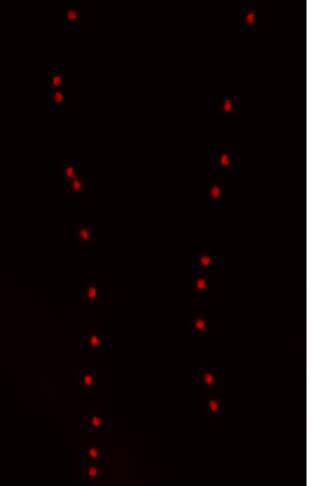
Brightfield



McrA-GFP

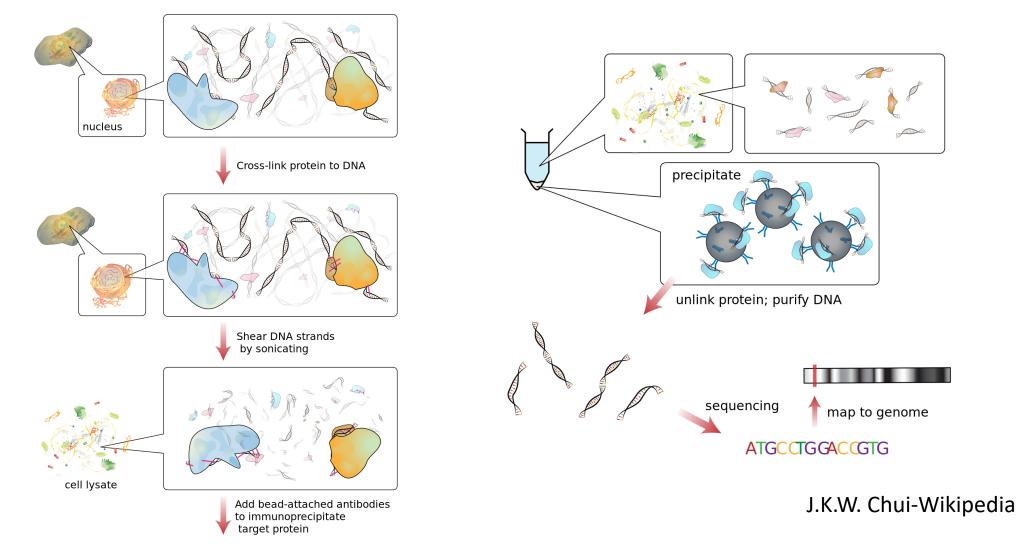


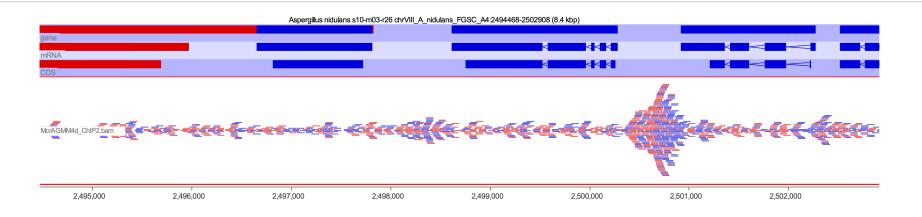
Histone H1 mCherry



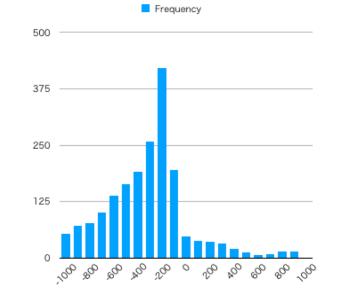


#### **ChIP-seq with HA-tagged McrA.**





#### 1899 statistically significant McrA ChIP-seq sites.



Distribution relative to transcription start sites. Most sites are in promoter regions of genes.



# Does McrA regulate secondary metabolism biosynthetic genes directly?

#### *McrA*<sup>Δ</sup> dramatically up-regulates expression of two nidulanin biosynthesis genes.

Gene	WT	mcrA∆	P<0.01?
nlsA (AN1242)	47.83 ± 35.08	447.26 ± 149.63	Yes
AN11934	38.86 ± 25.10	340.97 ± 67.61	Yes

Glucose minimal medium 4 days incubation. All values are reads per million, means +/- standard deviations, three biological replicates. Incubation at 37°C. P value from EdgeR analysis.

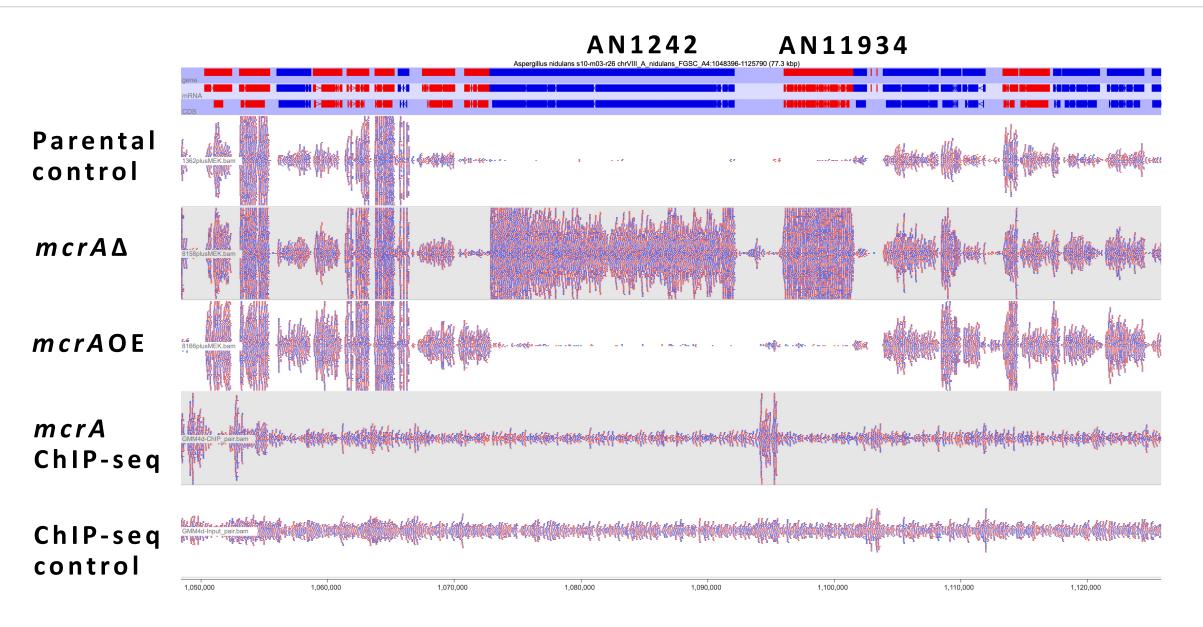


# **Over-expression of McrA dramatically down-regulates expression of nidulanin biosynthesis genes.**

Wild-type			alcA(p)-mcrA			
Gene	Uninduced	Induced	P<0.01?	Uninduced	Induced	P<0.01?
nlsA(AN1242)	72.81 ± 38.07	35.94 ± 14.43	No	142.38 ± 21.13	7.13 ± 1.92	Yes
AN11934	93.75 ± 14.23	102.75 ± 28.12	No	104.03 ± 13.51	13.57 ± 4.87	Yes

Lactose minimal medium, 3.5 days ± 12 h 50 mM methyl-ethyl ketone. All values reads per million, means ± standard deviation, three biological replicates. Incubation at 37°C.







Five secondary metabolism gene clusters have McrA ChIP-seq sites upstream of every gene and are regulated by McrA:

- Nidulanins
- F9775A,B/Orsellinic Acid
- Emericellamides (Production of the compounds is downregulated by McrAOE. McrA deletion does not substantially up-regulate.)
- AN10886-AN12290 (de-repressed, transcript levels moderate)
- AN9005-AN9006 (de-repressed, transcript levels moderate)



Many secondary metabolism gene clusters contain transcription factors that can, in principle, drive expression of the other genes of the cluster.

Does McrA regulate secondary metabolism via regulation of secondary-metabolism-specific transcription factors?



## Monodictyphenone biosynthetic gene cluster. 4 days incubation on glucose minimal medium

Gene	P<0.01?	Fold increase in <i>mcrA</i> ∆	Mean RPM in <i>mcrA</i> ∆	McrA-ChIP?	Predicted function of protein
AN10023/mdpL	Yes	165.03	1203.11		Member of the monodictyphenone (mdp) secondary metabolite biosynthesis gene cluster
AN10044/mdpK	Yes	183.24	1011.36		Putative oxidoreductase
AN10038/mdpJ	Yes	193.85	1275.62		Putative glutathione S transferase
AN10035/mdpl	Yes	186.96	165.54		Putative AMP-binding CoA ligase
AN10022/mdpH	Yes	148.73	1217.80		Protein with homology to the DUF 1772 superfamily
AN0150/mdpG	Yes	107.03	1447.14		Polyketide synthase
AN0149/mdpF	Yes	282.69	1415.20		Putative zinc-dependent hydrolase
AN0148/mdpE	Yes	188.68	232.86	Yes	C6 zinc finger transcription factor similar to AflR
AN0147/mdpD	Yes	146.15	308.30		Flavin-containing monooxygenase
AN0146/mdpC	Yes	171.56	2605.77		Protein with homology to versicolorin ketoreductase
AN10049/mdpB	Yes	107.13	980.66		Protein with homology to scytalone dehydratase
AN10021/mdpA	Yes	93.26	153.55		Regulatory protein



- 14 secondary metabolism gene clusters have McrA ChIPseq sites upstream of a transcription factor.
- At least 6 are strongly upregulated by *mcrA*∆ under some conditions.
- Transcription of others may depend on the availability of an activating transcription factor, chromatin access, etc.



## **Does McrA regulate regulators?**

Yes

Grau, M. F., R. Entwistle, C. E. Oakley, C. C. C. Wang, and B. R. Oakley. 2019. Overexpression of an LaeA-like Methyltransferase Upregulates Secondary Metabolite Production in Aspergillus nidulans. ACS Chem Biol **14**:1643-1651.

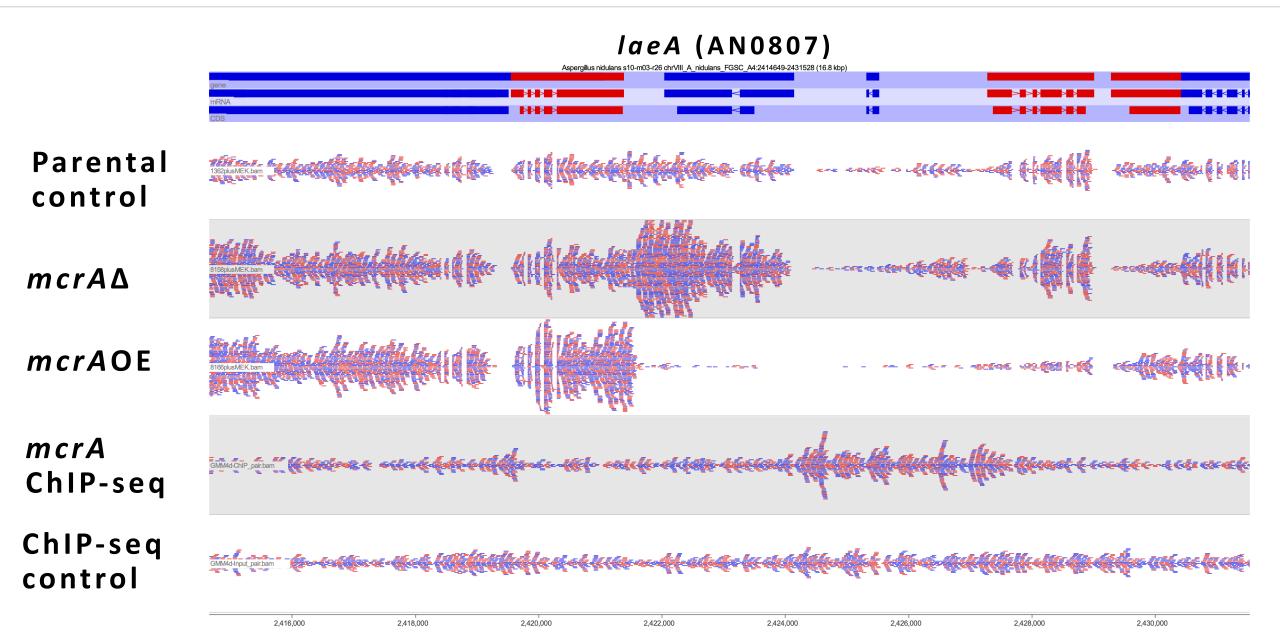
Gene	wt	mcrAΔ	p<0.01
<i>laeA</i> (AN0807)	20.77 +/- 4.05	37.84 +/- 11.18	no
<i>veA</i> (AN1052)	260.43 +/- 20.01	460.94 +/- 150.08	no
<i>velB</i> (AN0360)	25.98 +/- 6.44	51.93 +/- 1.52	no

## GMM (4days of growth)



	LMM <i>alcA</i> (p)- <i>mcrA</i> (3.5 days at 37ºC 12 h +/- 50 mM MEK)			
Gene	Uninduced	Induced	P<0.01?	
<i>laeA</i> (AN0807)	27.64 +/- 24.87	0.94 +/- 0.92	Yes	
<i>veA</i> (AN1052)	816.54 +/- 108.12	28.18 +/- 4.93	Yes	
<i>velB</i> (AN0360)	65.58 +/- 8.08	61.00 +/- 4.00	No	



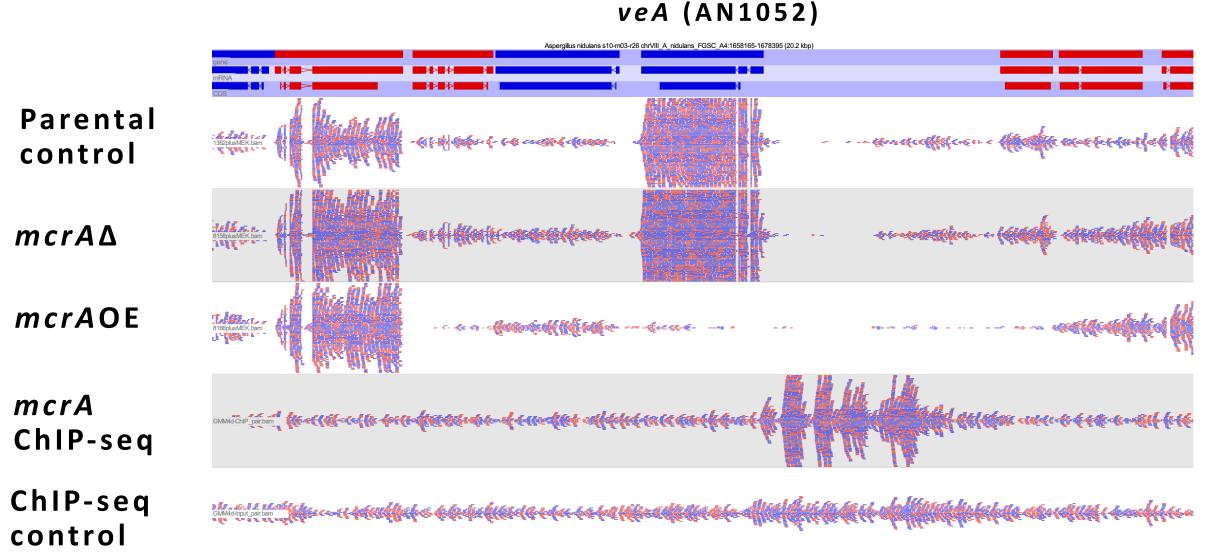


1,660,000

1.662.500

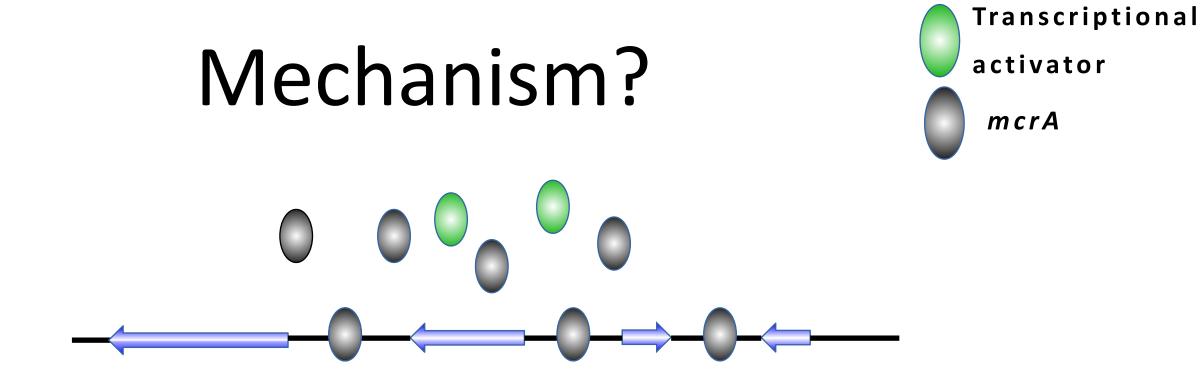
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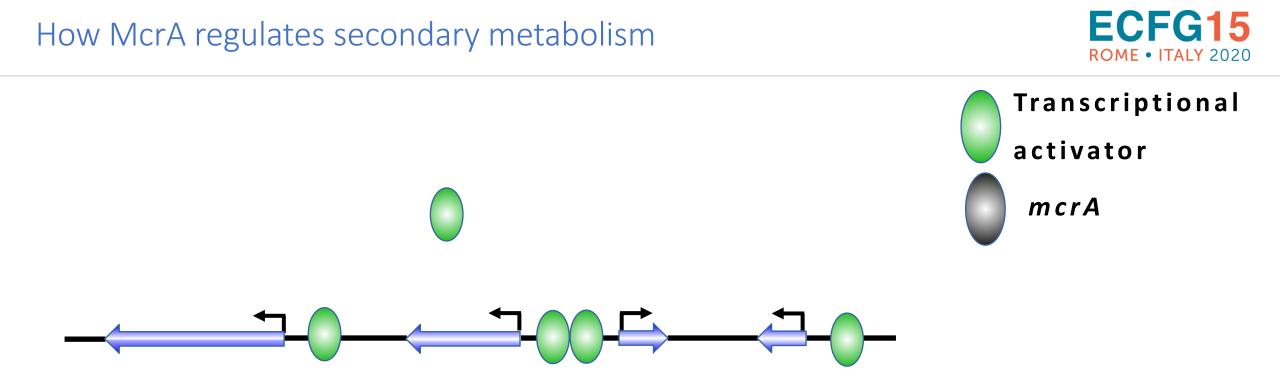


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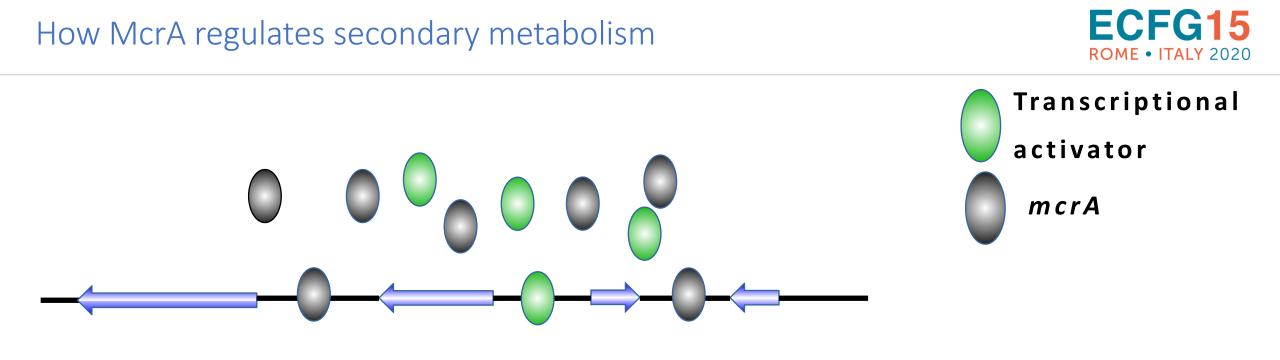




McrA binds to the promoter region of each gene and represses transcription.

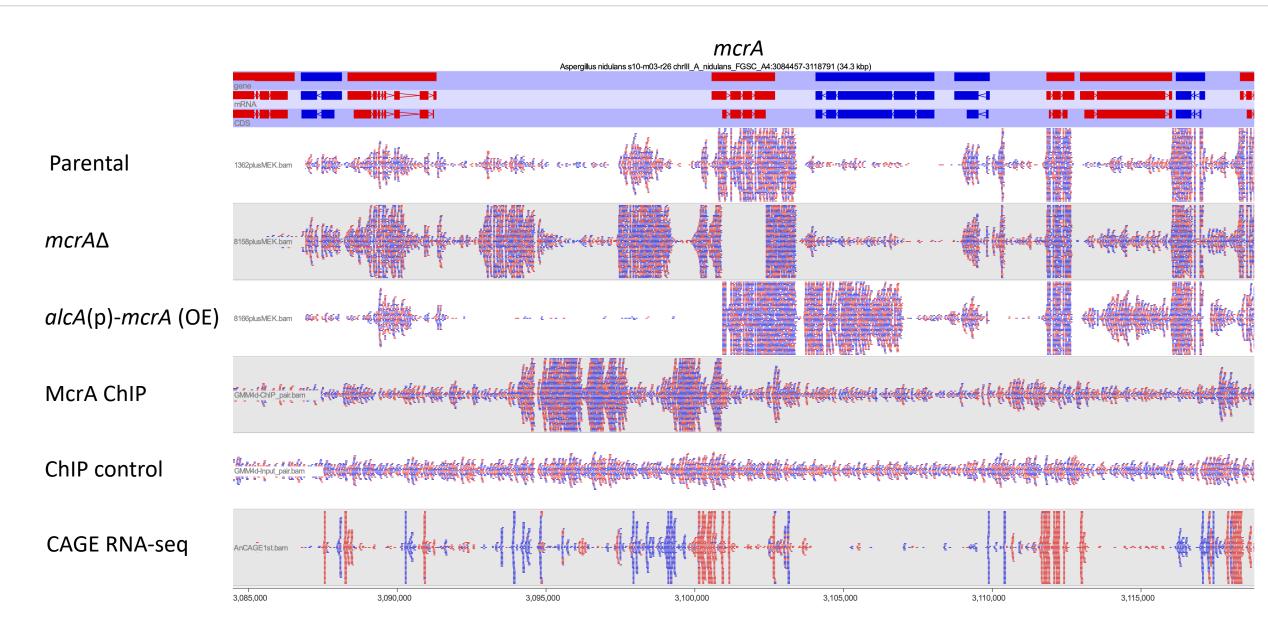


# Deletion of *mcrA* derepresses, but levels of transcription depend on the amount and activity of the transcriptional activator(s).



In the normal (*mcrA*+) situation, McrA and transcriptional activators are in equilibrium. Transcription depends on the relative abundances and activities of McrA and transcriptional activators (including affinities for binding sites).







- McrA binds upstream of many genes involved in secondary metabolism, likely functioning as a competitive inhibitor of transcription.
- It can regulate secondary metabolism genes directly and through cluster-specific transcription factors.
- It plays a role in regulating other secondary metabolism regulators.
- The *mcrA* region is transcriptionally complex and *mcrA* may autoregulate through a negative feedback loop.



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